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=> file chemistry	SINCE FILE	TOTAL
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=> polysaccharide
=> s polysaccharide
 40 FILES SEARCHED...
L1 239378 POLYSACCHARIDE

=> s li and glycosaminoglycan
 39 FILES SEARCHED...
L2 40 LI AND GLYCOSAMINOGLYCAN

=> s .12 and K5
L3 1 L2 AND K5

=> dis l3 bib abs

L3 ANSWER 1 OF 1 SCISEARCH COPYRIGHT 2002 ISI (R)
AN 97:816986 SCISEARCH
GA The Genuine Article (R) Number: YD473
TI Biosynthesis of heparin/heparan sulfate - DNA cloning and expression of
 D-glucuronyl C5-epimerase from bovine lung
AU Li J P (Reprint); HagnerMcWhirter A; Kjellen L; Palgi J; Jalalanen M;
 Lindahl U
CS UNIV UPPSALA, DEPT MED & PHYSIOL CHEM, POB 575, S-75123 UPPSALA, SWEDEN
 (Reprint); SWEDISH UNIV AGR SCI, CTR BIOMED, DEPT VET MED CHEM, S-75123
 UPPSALA, SWEDEN; UNIV TURKU, CTR BIOTECHNOL, FIN-20520 TURKU, FINLAND; ABO
 AKAD UNIV, FIN-20520 TURKU, FINLAND
CYA SWEDEN; FINLAND
SO JOURNAL OF BIOLOGICAL CHEMISTRY, (31 OCT 1997) Vol. 272, No. 44, pp.
 28158-28163.
 Publisher: AMER SOC BIOCHEMISTRY MOLECULAR BIOLOGY INC, 9650 ROCKVILLE

PIKE, BETHESDA, MD 20814.
ISSN: 0021-9258.

DT Article; Journal

FS LIFE

LA English

REC Reference Count: 33

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB Glucuronyl C5-epimerases catalyze the conversion of D-glucuronic acid (GlcUA) to L-iduronic acid (IdceA) units during the biosynthesis of **glycosaminoglycans**. An epimerase implicated in the generation of heparin/heparan sulfate was previously purified to homogeneity from bovine liver (Campbell, P., Hannesson, H. H., Sandback, D., Roden, L., Lindahl, U., and Li, J.-p, (1994) J. Biol. Chem. 269, 2695-26958). The present report describes the molecular cloning and functional expression of the lung enzyme. The cloned enzyme contains 444 amino acid residues and has a molecular mass of 49,905 Da. N-terminal sequence analysis of the isolated liver enzyme showed this species to be a truncated form lacking a 73-residue N-terminal domain of the deduced amino acid sequence.

The coding cDNA insert was cloned into a baculovirus expression vector and expressed in Sf9 insect cells. Cells infected with recombinant epimerase showed a 20-30-fold increase in enzyme activity, measured as release of (H₂O)-H-3 from a polysaccharide substrate containing C5-H-3-labeled hexuronic acid units. Furthermore, incubation of the expressed protein with the appropriate (GlcUA-GlcNSO(3))(n) substrate resulted in conversion of similar to 20% of the GlcUA units into IdceA residues. Northern analysis implicated two epimerase transcripts in both bovine lung and liver tissues, a dominant similar to 9-kilobase (kb) mRNA and a minor similar to 5-kb species. Mouse mastocytoma cells showed only the similar to 5-kb transcript. A comparison of the cloned epimerase with the enzymes catalyzing an analogous reaction in alginate biosynthesis revealed no apparent amino acid sequence similarity.

=> s 11 and K5
42 FILES SEARCHED...
L4 351 L1 AND K5

=> s 14 and epimer?
L5 33 L4 AND EPIMER?

=> s 15 and enzyme
36 FILES SEARCHED...
L6 20 L5 AND ENZYME

=> s 16 and immobil?
37 FILES SEARCHED...
L7 5 L6 AND IMMOBIL?

=> dis 17 1-5 bib abs

L7 ANSWER 1 OF 5 BIOTECHNO COPYRIGHT 2002 Elsevier Science B.V.
AN 1994:24332896 BIOTECHNO
TI Biosynthesis of heparin/heparan sulfate. Purification of the D-glucuronyl C-5 **epimerase** from bovine liver
AU Campbell P.; Hannesson H.H.; Sandback D.; Roden L.; Lindahl U.; Li J.-P.
CS Dept. of Medical/Physiological Chem., Biomedical Center, University of Uppsala, S-751 23 Uppsala, Sweden.
SO Journal of Biological Chemistry, (1994), 269/43 (26953-26958)
CODEN: JBCHA3 ISSN: 0021-9258
DT Journal; Article
CY United States
LA English
SL English
AB The D-glucuronyl C-5 **epimerase** involved in the biosynthesis of

heparin/heparan sulfate was purified from the high speed supernatant fraction of a homogenate of bovine liver by chromatography on **immobilized** O-desulfated heparin, red Sepharose, phenyl Sepharose, and concanavalin A-Sepharose. After close to 1 million-fold purification, in 10-15% yield, the product gave a single band on SDS-polyacrylamide gel electrophoresis with silver staining and had a mobility corresponding to an M(r) of .sim.52,000. Since the **epimerase** assay used in the course of purification was based on release of tritium, as .cents..sup.3H!H.sub.2O, from a .cents.5-.sup.3H!uronyl-labeled substrate, it was important to establish that the purified **enzyme** did indeed catalyze the actual conversion of D-glucuronyl to L-iduronyl residues. Upon incubation of the purified **enzyme** with .sup.3H-labeled heparosan N-sulfate, prepared by metabolic labeling (with D-.cents.1-.sup.3H!glucose) of a capsular **polysaccharide** from Escherichia coli **K5** and subsequent chemical partial N-deacetylation and N-sulfation, approximately 30% of the D-glucuronyl residues located between two N-sulfated glucosamine units were converted to L-iduronyl units.

L7 ANSWER 2 OF 5 CAPLUS COPYRIGHT 2002 ACS
AN 2002:392262 CAPLUS
DN 136:380105
TI Glycosaminoglycans derived from **k5 polysaccharide**
having high anticoagulant and antithrombotic activities and process for
their preparation
IN Oreste, Pasqua; Zoppetti, Giorgio
PA Italy
SO U.S. Pat. Appl. Publ., 39 pp., Cont.-in-part of U.S. Ser. No. 738,879.
CODEN: USXXCO
DT Patent
LA English
FAN.CNT 2

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 2002062019	A1	20020523	US 2001-950003	20010912
	IT 2000MI0665	A1	20011001	IT 2000-MI665	20000330
	WO 2002050125	A2	20020627	WO 2001-IB2492	20011217
	WO 2002050125	A3	20020822		
		W:	AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM		
		RW:	GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG		
PRAI	IT 2000-MI665	A	20000330		
	US 2000-738879	A2	20001218		
	US 2001-950003	A	20010912		
AB	Glycosaminoglycans derived from K5 polysaccharide having high anticoagulant and antithrombotic activity and useful for the control of coagulation and as antithrombotic agents are obtained starting from an optionally purified K5 polysaccharide by a process comprising the steps of N-deacetylation/N-sulfation, C5 epimerization , O-versulfation, selective O-desulfation, 6-O-sulfation, N-sulfation, and optional depolymerization, in which said epimerization is performed with the use of the enzyme glucoronosyl C5 epimerase in soln. or in immobilized form in the presence of divalent cations. New, particularly interesting antithrombin compds. are obtained by controlling the reaction time in the selective O-desulfation step and submitting the product obtained at the end of the final N-sulfation step to depolymerization.				

L7 ANSWER 3 OF 5 CAPLUS COPYRIGHT 2002 ACS
 AN 2001:730838 CAPLUS
 DN 135:267246
 TI Glycosaminoglycans derived from the **k5 polysaccharide**
 having high anticoagulant and antithrombotic activity and process for
 their preparation
 IN Zoppetti, Giorgio; Oreste, Pasqua; Cipolletti, Giovanni
 PA Inalco S.p.A., Italy
 SO PCT Int. Appl., 38 pp.
 CODEN: PIXXD2
 DT Patent
 LA English
 FAN.CNT 2

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2001072848	A1	20011004	WO 2001-EP3461	20010327
	W:	AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM			
	RW:	GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG			
	IT 2000MI0665	A1	20011001	IT 2000-MI665	20000330
PRAI	IT 2000-MI665	A	20000330		

AB Glycosaminoglycans derived from the **K5 polysaccharide**
 having high anticoagulant and antithrombotic activity obtained by a
 process comprising the prepn. of the **K5 polysaccharide**
 from *Escherichia coli*, N-deacetylation/N-sulfation, C-5
epimerization, supersulfation, selective O-desulfation, selective
6-O sulfation and N-sulfation, wherein said **epimerization** is
carried out using the glucuronosyl C-5 **epimerase enzyme**
in soln. or in **immobilized** form in presence of specific divalent
cations.

RE.CNT 4 THERE ARE 4 CITED REFERENCES AVAILABLE FOR THIS RECORD
 ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 4 OF 5 CAPLUS COPYRIGHT 2002 ACS
 AN 1994:624763 CAPLUS
 DN 121:224763
 TI Biosynthesis of heparin/heparan sulfate. Purification of the D-glucuronyl
C-5 **epimerase** from bovine liver
 AU Campbell, Patrick; Hannesson, Helgi H.; Sandbaek, Dagmar; Roden,
Lennart; Lindahl, Ulf; Li, Jin-ping
 CS Univ. Alabama, Birmingham, AL, 35294, USA
 SO Journal of Biological Chemistry (1994), 269(43), 26953-8
 CODEN: JBCHA3; ISSN: 0021-9258
 DT Journal
 LA English
 AB The D-glucuronyl C-5 **epimerase** involved in the biosynthesis of
heparin/heparan sulfate was purified from the high speed supernatant
fraction of a homogenate of bovine liver by chromatog. on
immobilized O-desulfate heparin, red Sepharose, Ph Sepharose, and
Con A-Sepharose. After close to 1 million-fold purifn., 10-15% yield, the
product gave a single band on SDS-PAGE with silver staining and had a
mobility corresponding to an Mr of .apprx.52,000. Since the
epimerase assay used in the course of purifn. was based on release
of tritium, as [3H]H₂O, from a [5-3H]uronyl-labeled substrate, it was
important to establish that the purified **enzyme** did indeed
catalyze the actual conversion of D-glucuronyl to L-iduronyl residues.
Upon incubation of the purified **enzyme** with 3H-labeled heparosan

N-sulfate, prep'd. by metabolic labeling (with D-[1-3H]glucose) of a capsular polysaccharide from Escherichia coli K5 and subsequent chem. partial N-deacetylation and N-sulfation, approx. 30% of the D-glucuronyl residues located between two N-sulfated glucosamine units were converted to L-iduronyl units.

L7 ANSWER 5 OF 5 SCISEARCH COPYRIGHT 2002 ISI (R)
AN 94:733790 SCISEARCH
GA The Genuine Article (R) Number: PQ931
TI BIOSYNTHESIS OF HEPARIN/HEPARAN SULFATE - PURIFICATION OF THE D-GLUCURONYL C-5 EPIMERASE FROM BOVINE LIVER
AU CAMPBELL P; HANNESSON H H; SANDBACK D; RODEN L; LINDAHL U; LI J P
(Reprint)
CS UNIV UPPSALA, DEPT MED & PHYSIOL CHEM, BOX 575, S-75123 UPPSALA, SWEDEN
(Reprint); UNIV UPPSALA, DEPT MED & PHYSIOL CHEM, S-75123 UPPSALA, SWEDEN;
UNIV ALABAMA, BIRMINGHAM, AL, 35294
CYA SWEDEN; USA
SO JOURNAL OF BIOLOGICAL CHEMISTRY, (28 OCT 1994) Vol. 269, No. 43, pp.
26953-26958.
ISSN: 0021-9258.

DT Article; Journal

FS LIFE

LA ENGLISH

REC Reference Count: 21

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB The D-glucuronyl C-5 epimerase involved in the biosynthesis of heparin/heparan sulfate was purified from the high speed supernatant fraction of a homogenate of bovine liver by chromatography on immobilized O-desulfated heparin, red Sepharose, phenyl Sepharose, and concanavalin A-Sepharose. After close to 1 million-fold purification, in 10-15% yield, the product gave a single band on SDS-polyacrylamide gel electrophoresis with silver staining and had a mobility corresponding to an M(r) of similar to 52,000. Since the epimerase assay used in the course of purification was based on release of tritium, as [H-3]H₂O, from a [5-H-3]uronyl-labeled substrate, it was important to establish that the purified enzyme did indeed catalyze the actual conversion of D-glucuronyl to L-iduronyl residues. Upon incubation of the purified enzyme with H-3-labeled heparosan N-sulfate, prepared by metabolic labeling (with D-[1-H-3]glucose) of a capsular polysaccharide from Escherichia coli K5 and subsequent chemical partial N-deacetylation and N-sulfation, approximately 30% of the D-glucuronyl residues located between two N-sulfated glucosamine units were converted to L-iduronyl units.

=> dis.hist

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L1 239378 S POLYSACCHARIDE
L2 40 S LI AND GLYCOSAMINOGLYCAN
L3 1 S L2 AND K5
L4 351 S L1 AND K5
L5 33 S L4 AND EPIMER?
L6 20 S L5 AND ENZYME
L7 5 S L6 AND IMMOBIL?

=> s 12 and composition
35 FILES SEARCHED...

L8 1 L2 AND COMPOSITION

=> dis 18 bib abs

L8 ANSWER 1 OF 1 CAPLUS COPYRIGHT 2002 ACS
AN 2002:532818 CAPLUS
TI Extracellular matrix **composition** of full-thickness defect repair tissue is little influenced by exercise in rat articular cartilage
AU Espanha, M. Margarida; Lammi, Pirkko E.; Hyttinen, Mika M.; Lammi, Mikko J.; Helminen, Heikki J.
CS Faculty of Human Kinetics, Technical University of Lisboa, Lisbon, Port.
SO Connective Tissue Research (2001), 42(2), 97-109
CODEN: CVTRBC; ISSN: 0300-8207
PB Gordon & Breach Science Publishers
DT Journal
LA English
AB Full-thickness articular cartilage defects in the femoral condyles of adult rats were examd. four and eight weeks after injury. Quant. polarized light microscopic anal. showed that birefringence of the tissue in the central repair area increased more in rats exercised on a treadmill. **Glycosaminoglycan** content in the repair tissue was also higher than in the intermittent active motion group at four weeks after injury, but by eight weeks the levels were similar in both groups. No normal-looking articular cartilage was formed in the lesions, and only in one animal type **II** collagen was obsd. in the superficial zone of repair tissue. No 3B3(-) antigenicity of the proteoglycans was seen during repair. In conclusion, exercise minimally modified the repair of full-thickness articular cartilage defects in adult rats. The repair in the exercised group may occur slightly faster in the early stages but no difference was seen at the eight week time interval between the exercised and the intermittently active group.

RE.CNT 56 THERE ARE 56 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

=> dis hist

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L1 239378 S POLYSACCHARIDE
L2 40 S L1 AND GLYCOSAMINOGLYCAN
L3 1 S L2 AND K5
L4 351 S L1 AND K5
L5 33 S L4 AND EPIMER?
L6 20 S L5 AND ENZYME
L7 5 S L6 AND IMMOBIL?
L8 1 S L2 AND COMPOSITION

=> s 14 and composition

39 FILES SEARCHED...

L9 13 L4 AND COMPOSITION

=> s 19 and glycosaminoglycan

42 FILES SEARCHED...

L10 0 L9 AND GLYCOSAMINOGLYCAN

=> s 19 and carrier

39 FILES SEARCHED...

L11 0 L9 AND CARRIER

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=> s 19 and ion?  
 20 FILES SEARCHED...  
 36 FILES SEARCHED...  
L12      1 L9 AND ION?
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=> dis l12 bib abs
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L12  ANSWER 1 OF 1  CAPLUS  COPYRIGHT 2002 ACS  
AN  1984:2544  CAPLUS  
DN  100:2544  
TI  Assay of N-acetylheparosan deacetylase with a capsular  
    polysaccharide from Escherichia coli K5 as substrate  
AU  Navia, Juan Luis; Riesenfeld, Johan; Vann, Willie F.; Lindahl, Ulf; Roden,  
    Lennart  
CS  Inst. Dent. Res., Univ. Alabama, Birmingham, AL, 35294, USA  
SO  Anal. Biochem. (1983), 135(1), 134-40  
    CODEN: ANBCA2; ISSN: 0003-2697  
DT  Journal  
LA  English  
AB  A new substrate for N-acetylheparosan deacetylase was prep'd. The capsular  
    polysaccharide from E. coli O10:K5:H4, which is  
    structurally identical to N-acetylheparosan, was partially N-deacetylated  
    by hydrazinolysis and was then radioactively labeled by N-acetylation with  
    [3H]acetic anhydride. Upon incubation of the labeled  
    polysaccharide with microsomes from the Furth mastocytoma,  
    [3H]acetyl groups were released, demonstrating that the bacterial  
    polysaccharide was a substrate for the N-deacetylase. Reaction  
    conditions were established which permitted the quant. assay of  
    N-deacetylase activity; a Km of 74 mg polysaccharide/L was detd.  
    which corresponds to 2.1 .times. 10-4M, expressed as concn. of uronic  
    acid; the Vmax was 3.4 nmol/mg protein/L. In confirmation of previous  
    results, it was obsd. (1) that the reaction was stimulated by  
    3'-phosphoadenylylsulfate (up to a max. of 45% at a concn. of 0.5 mM),  
    suggesting that N-sulfation occurred which facilitated continued action of  
    the N-deacetylase, and (2) that NaCl and KCl inhibited the enzyme, with  
    50% redn. of activity at a concn. of 25 mM. In the course of this work, a  
    simple, single-vial assay procedure was used. Released [3H]acetate was  
    extd. from the acidified reaction mixt. with a toluene- or xylene-based  
    scintillation fluid contg. 10% isoamyl alc. and measured directly by  
    scintillation spectrometry.
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```
=> s glycosaminoglycans  
L13      23302 GLYCOSAMINOGLYCANs
```

```
=> s l13 and K5  
L14      47 L13 AND K5
```

```
=> s l14 and process  
  25 FILES SEARCHED...  
L15      7 L14 AND PROCESS
```

```
=> s l15 deacetyla?  
MISSING OPERATOR L15 DEACETYLA?  
The search profile that was entered contains terms or  
nested terms that are not separated by a logical operator.
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```
=> s l15 and deacetyla?  
L16      6 L15 AND DEACETYLA?
```

```
=> s l16 and N-sulfat?  
75% OF LIMIT FOR L#S REACHED  
  26 FILES SEARCHED...
```

L17 6 L16 AND N-SULFAT?

=> dis 117 1-6 bib abs

L17 ANSWER 1 OF 6 BIOTECHNO COPYRIGHT 2002 Elsevier Science B.V.
AN 2001:32989368 BIOTECHNO
TI Toward a biotechnological heparin through combined chemical and enzymatic
modification of the Escherichia coli K5 polysaccharide
AU Naggi A.; Torri G.; Casu B.; Oreste P.; Zoppetti G.; Li J.P.; Lindahl U.
CS Prof. B. Casu, G. Ronzoni Research Institute, via G. Colombo, 81-20133
Milano, Italy.
E-mail: casu@ronzoni.it
SO Seminars in Thrombosis and Hemostasis, (2001), 27/5 (437-443), 26
reference(s)
CODEN: STHMBV ISSN: 0094-6176
DT Journal; General Review
CY United States
LA English
SL English
AB A **process** to generate **glycosaminoglycans** with
heparin- and heparan sulfate-like sequences from the Escherichia coli
K5 capsular polysaccharide is described. This polymer has the
same structure as N-acetylheparosan, the precursor in heparin/heparan
sulfate biosynthesis. The **process** involves chemical N-
deacetylation and **N-sulfation**, enzymatic,
conversion of up to 60% of the D-glucuronic acid to L-iduronic acid
residues, and chemical O-sulfation. Because direct sulfation afforded
unwanted 3-O-sulfated (instead of 2-O-sulfated) iduronic acid residues, a
strategy involving graded solvolytic desulfation of chemically
oversulfated C5-epimerized sulfaminoheparosans was assessed using
persulfated heparin and heparan sulfate as model compounds. The
O-desulfation **process** was shown to increase the anti-factor Xa
activity of oversulfated heparin.

L17 ANSWER 2 OF 6 CAPLUS COPYRIGHT 2002 ACS
AN 2002:392262 CAPLUS
DN 136:380105
TI **Glycosaminoglycans** derived from **k5** polysaccharide
having high anticoagulant and antithrombotic activities and
process for their preparation
IN Oreste, Pasqua; Zoppetti, Giorgio
PA Italy
SO U.S. Pat. Appl. Publ., 39 pp., Cont.-in-part of U.S. Ser. No. 738,879.
CODEN: USXXCO
DT Patent
LA English
FAN.CNT 2

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 2002062019	A1	20020523	US 2001-950003	20010912
	IT 2000MI0665	A1	20011001	IT 2000-MI665	20000330
	WO 2002050125	A2	20020627	WO 2001-IB2492	20011217
	WO 2002050125	A3	20020822		
				W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GO, GW, ML, MR, NE, SN, TD, TG	
	PRAI IT 2000-MI665	A	20000330		

US 2000-738879 A2 20001218
US 2001-950003 A 20010912

AB **Glycosaminoglycans** derived from **K5** polysaccharide having high anticoagulant and antithrombotic activity and useful for the control of coagulation and as antithrombotic agents are obtained starting from an optionally purified **K5** polysaccharide by a process comprising the steps of **N-deacetylation/N-sulfation**, C5 epimerization, O-versulfation, selective O-desulfation, 6-O-sulfation, **N-sulfation**, and optional depolymerization, in which said epimerization is performed with the use of the enzyme glucoronosyl C5 epimerase in soln. or in immobilized form in the presence of divalent cations. New, particularly interesting antithrombin compds. are obtained by controlling the reaction time in the selective O-desulfation step and submitting the product obtained at the end of the final **N-sulfation** step to depolymerization.

L17 ANSWER 3 OF 6 CAPLUS COPYRIGHT 2002 ACS
AN 2001:840398 CAPLUS
DN 136:167613
TI Toward a biotechnological heparin through combined chemical and enzymatic modification of the Escherichia coli **K5** polysaccharide
AU Naggi, Annamaria; Torri, Giangiacomo; Casu, Benito; Oreste, Pasqua; Zoppetti, Giorgio; Li, Jin-Ping; Lindahl, Ulf
CS G. Ronzoni Institute for Chemical and Biochemical Research, Milan, Italy
SO Seminars in Thrombosis and Hemostasis (2001), 27(5), 437-443 CODEN: STHMBV; ISSN: 0094-6176
PB Thieme Medical Publishers, Inc.
DT Journal
LA English
AB A process to generate **glycosaminoglycans** with heparin and heparan sulfate-like sequences from the Escherichia coli **K5** capsular polysaccharide is described. This polymer has the same structure as N-acetylheparosan, the precursor in heparin/heparan sulfate biosynthesis. The process involves chem. N-deacetylation and N-sulfation, enzymic conversion of up to 60% of the D-glucuronic acid to L-iduronic acid residues, and chem. O-sulfation. Because direct sulfation afforded unwanted 3-O-sulfated (instead of 2-O-sulfated) iduronic acid residues, a strategy involving graded solvolytic desulfation of chem. over-sulfated C5-epimerized sulfaminoheparosans was assessed using persulfated heparin and heparan sulfate as model compds. The O-desulfation process was shown to increase the anti-factor Xa activity of over-sulfated heparin.

RE.CNT 26 THERE ARE 26 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L17 ANSWER 4 OF 6 CAPLUS COPYRIGHT 2002 ACS
AN 2001:730838 CAPLUS
DN 135:267246
TI **Glycosaminoglycans** derived from the **k5** polysaccharide having high anticoagulant and antithrombotic activity and process for their preparation
IN Zoppetti, Giorgio; Oreste, Pasqua; Cipolletti, Giovanni
PA Inalco S.p.A., Italy
SO PCT Int. Appl., 38 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 2

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
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PI WO 2001072848	A1	20011004	WO 2001-EP3461	20010327
W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM,				

HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS,
LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO,
RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ,
VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM
RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY,
DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF,
BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG

IT 2000MI0665 A1 20011001 IT 2000-MI665 20000330

PRAI IT 2000-MI665 A 20000330

AB **Glycosaminoglycans** derived from the **K5** polysaccharide having high anticoagulant and antithrombotic activity obtained by a process comprising the prepn. of the **K5** polysaccharide from *Escherichia coli*, **N-deacetylation/N-sulfation**, C-5 epimerization, supersulfation, selective O-desulfation, selective 6-O sulfation and **N-sulfation**, wherein said epimerization is carried out using the glucuronosyl C-5 epimerase enzyme in soln. or in immobilized form in presence of specific divalent cations.

RE.CNT 4 THERE ARE 4 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L17 ANSWER 5 OF 6 PASCAL COPYRIGHT 2002 INIST-CNRS. ALL RIGHTS RESERVED.
AN 2002-0168335 PASCAL
CP Copyright .COPYRGT. 2002 INIST-CNRS. All rights reserved.
TIEN Toward a biotechnological heparin through combined chemical and enzymatic modification of the *Escherichia coli* **K5** polysaccharide Glycoaminoglycans
AU NAGGI Annamaria; TORRI Giangiacomo; CASU Benito; ORESTE Pasqua; ZOPPETTI Giorgio; LI Jin-Ping; LINDAHL Ulf HARENBERG Job (ed.)
CS G. Ronzoni Institute for Chemical and Biochemical Research, Milan, Italy; Inalco, Montale, Pistoia, Italy; Department of Medical Biochemical Microbiology, University of Uppsala, Uppsala, Sweden
SO Seminars in thrombosis and hemostasis, (2001), 27(5), 437-443, 26 refs.
ISSN: 0094-6176 CODEN: STHMBV
DT Journal
BL Analytic
CY United States
LA English
AV INIST-17786, 354000094741730010
CP Copyright .COPYRGT. 2002 INIST-CNRS. All rights reserved.
AB A process to generate **glycosaminoglycans** with heparin- and heparan sulfate-like sequences from the *Escherichia coli* **K5** capsular polysaccharide is described. This polymer has the same structure as N-acetylheparosan, the precursor in heparin/ heparan sulfate biosynthesis. The process involves chemical **N-deacetylation** and **N-sulfation**, enzymatic conversion of up to 60% of the D-glucuronic acid to L-iduronic acid residues, and chemical O-sulfation. Because direct sulfation afforded unwanted 3-O-sulfated (instead of 2-O-sulfated) iduronic acid residues, a strategy involving graded solvolytic desulfation of chemically oversulfated C5-epimerized sulfaminoheparosans was assessed using persulfated heparin and heparan sulfate as model compounds. The O-desulfation process was shown to increase the anti-factor Xa activity of oversulfated heparin.

L17 ANSWER 6 OF 6 SCISEARCH COPYRIGHT 2002 ISI (R)
AN 2001:876964 SCISEARCH
GA The Genuine Article (R) Number: 486YG
TI Toward a biotechnological heparin through combined chemical and enzymatic modification of the *Escherichia coli* **K5** polysaccharide
AU Naggi A; Torri G; Casu B (Reprint); Oreste P; Zoppetti G; Li J P; Lindahl U
CS G Ronzoni Res Inst, Via G Colombo, 81, I-20133 Milan, Italy (Reprint); G

Ronzoni Res Inst, I-20133 Milan, Italy; G Ronzoni Inst Chem & Biochem Res, Milan, Italy; Inalco, Montale, Pistoia, Italy; Univ Uppsala, Dept Med Biochem Microbiol, S-75105 Uppsala, Sweden
CYA Italy; Sweden ^{in D}
SO SEMINARS IN THROMBOSIS AND HEMOSTASIS, (OCT 2001) Vol. 27, No. 5, pp.
437-443.
Publisher: THIEME MEDICAL PUBL INC, 333 SEVENTH AVE, NEW YORK, NY 10001 USA.
ISSN: 0094-6176.
DT Article; Journal
LA English
REC Reference Count: 26
ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS
AB A **process** to generate **glycosaminoglycans** with heparin- and heparan sulfate-like sequences from the Escherichia coli K5 capsular polysaccharide is described. This polymer has the same structure as N-acetylheparosan, the precursor in heparin/ heparan sulfate biosynthesis. The **process** involves chemical N-deacetylation and N-sulfation, enzymatic conversion of up to 60% of the D-glucuronic acid to L-iduronic acid residues, and chemical O-sulfation. Because direct sulfation afforded unwanted 3-O-sulfated (instead of 2-O-sulfated) iduronic acid residues, a strategy involving graded solvolytic desulfation of chemically oversulfated C5-epimerized sulfaminoheparosans was assessed using persulfated heparin and heparan sulfate as model compounds. The O-desulfation **process** was shown to increase the anti-factor Xa activity of oversulfated heparin.